3. If the above method does not succeed, allow the small pieces of fresh tissue to remain exposed to the action of the air for ten to twenty minutes before placing in the Flemming solution. This should be at a temperature of 4° C. Time as in 2.

4. Wash in water about 24 hours.

5. Dehydrate by very gradual steps.

6. Clear from 95 per cent alcohol in cedar oil followed by xylol.

7. Imbed in paraffin.

QUINCE JELLY AS A CULTURE MEDIUM FOR EUGLENA

Turner (Anat. Rec., 12:3, April, 1917) develops a mode of culturing Euglena and other green protozoa in quince jelly, which has practical value for the general laboratory because of the ease of making and preserving the cultures. Cultures have been kept for more than a year, and may be successfully transplanted.

The methods and results may be summarized as follows:

1. Twenty grams of dry quince seed boiled for half an hour in 1.5 liters of distilled water produces enough of the exudate, after the seeds are strained out, to make a total culture mass of 2½ liters, by the addition of distilled water.

2. A still more fruitful medium was made by grinding the seeds to a fine powder or meal and leaving it in the jelly. Water to 2½ or 3 liters may be used with this amount of meal. This medium has the disadvantage of being opaque.

3. Cultures develop more rapidly in a slightly thinner medium, but more lasting cultures call for a thicker medium.

4. The medium should be made slightly alkaline, and the cultures kept at room temperature, in moderate light.

5. Test tubes, vials, and similar vessels furnishing large vertical range and little exposed surface are best.

6. The reactions of Euglena are modified in interesting ways, as compared with their usual behavior in water.

"VITAL DYES" IN TELEOSTS

Wislocki (Anat. Record. XII, 4, May 1917) reports on experiments with benzidine dyes injected into the peritoneal cavity of